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APPLICATION THAT MET THE REQUIREMENTS TO BE GRANTED A
FILING DATE.

APPLICATION NUMBER: 60/532,681

FILING DATE: *December 23, 2003*

RELATED PCT APPLICATION NUMBER: PCT/US04/43499

Certified by



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Under Secretary of Commerce
for Intellectual Property
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PROVISIONAL APPLICATION FOR PATENT COVER SHEET

This is a request for filing a PROVISIONAL APPLICATION FOR PATENT under 37 CFR 1.53(c).

Express Mail Label No. EV 389269196

INVENTOR(S)

Given Name (first and middle [if any])	Family Name or Surname	Residence (City and either State or Foreign Country)
Vijay K. Song-Hua	Mahant Ke	Murrieta, CA San Diego, CA

Additional inventors are being named on the _____ separately numbered sheets attached hereto

TITLE OF THE INVENTION (500 characters max)

Biochip Configurations and Methods

Direct all correspondence to: **CORRESPONDENCE ADDRESS**

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Individual Name

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ENCLOSED APPLICATION PARTS (check all that apply)

Specification Number of Pages 5 CD(s), Number _____
 Drawing(s) Number of Sheets 1 Other (specify) _____
 Application Data Sheet. See 37 CFR 1.76

METHOD OF PAYMENT OF FILING FEES FOR THIS PROVISIONAL APPLICATION FOR PATENT

Applicant claims small entity status. See 37 CFR 1.27.
 A check or money order is enclosed to cover the filing fees.
 The Director is hereby authorized to charge filing fees or credit any overpayment to Deposit Account Number: 502191 FILING FEE Amount (\$)
80.00
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The invention was made by an agency of the United States Government or under a contract with an agency of the United States Government.

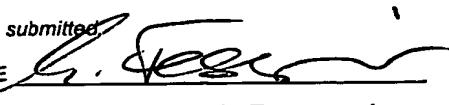
No.

Yes, the name of the U.S. Government agency and the Government contract number are: _____

[Page 1 of 1]

Date 12/23/03

Respectfully submitted

SIGNATURE 

TYPED or PRINTED NAME Martin Fessenmaier

TELEPHONE 714-641-5100

REGISTRATION NO. 46697

(If appropriate) Docket Number: 100788.0022PRO

USE ONLY FOR FILING A PROVISIONAL APPLICATION FOR PATENT
 This collection of information is required by 37 CFR 1.51. The information is required to obtain or retain a benefit by the public which is to file (and by the USPTO to process) an application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.14. This collection is estimated to take 8 hours to complete, including gathering, preparing, and submitting the completed application form to the USPTO. Time will vary depending upon the individual case. Any comments on the amount of time you require to complete this form and/or suggestions for reducing this burden should be sent to the Chief Information Officer, U.S. Patent and Trademark Office, U.S. Department of Commerce, P.O. Box 1450, Alexandria, VA 22313-1450. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Mail Stop Provisional Application, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450.

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FEE TRANSMITTAL

for FY 2004

Effective 10/01/2003. Patent fees are subject to annual revision.

 Applicant claims small entity status. See 37 CFR 1.27

TOTAL AMOUNT OF PAYMENT (\$ 80.00)

Complete if Known

Application Number	
Filing Date	December 23, 2003
First Named Inventor	Vijay K. Mahant
Examiner Name	
Art Unit	
Attorney Docket No.	100788.0022PRO

METHOD OF PAYMENT (check all that apply)

 Check Credit card Money Order Other None
 Deposit Account:

Deposit Account Number	502191
Deposit Account Name	Rutan & Tucker

The Director is authorized to: (check all that apply)

 Charge fee(s) indicated below Credit any overpayments
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FEE CALCULATION

1. BASIC FILING FEE

Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid
1001 770	2001 385	Utility filing fee	
1002 340	2002 170	Design filing fee	
1003 530	2003 265	Plant filing fee	
1004 770	2004 385	Reissue filing fee	
1005 160	2005 80	Provisional filing fee	80.00
SUBTOTAL (1) (\$)			80.00

2. EXTRA CLAIM FEES FOR UTILITY AND REISSUE

Total Claims	Independent Claims	Extra Claims	Fee from below	Fee Paid
		-20** =	X	=
		- 3** =	X	=

Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description
1202 18	2202 9	Claims in excess of 20
1201 86	2201 43	Independent claims in excess of 3
1203 290	2203 145	Multiple dependent claim, if not paid
1204 86	2204 43	** Reissue independent claims over original patent
1205 18	2205 9	** Reissue claims in excess of 20 and over original patent
SUBTOTAL (2) (\$)		

**or number previously paid, if greater; For Reissues, see above

3. ADDITIONAL FEES

Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid
1051 130	2051 65	Surcharge - late filing fee or oath	
1052 50	2052 25	Surcharge - late provisional filing fee or cover sheet	
1053 130	1053 130	Non-English specification	
1812 2,520	1812 2,520	For filing a request for ex parte reexamination	
1804 920*	1804 920*	Requesting publication of SIR prior to Examiner action	
1805 1,840*	1805 1,840*	Requesting publication of SIR after Examiner action	
1251 110	2251 55	Extension for reply within first month	
1252 420	2252 210	Extension for reply within second month	
1253 950	2253 475	Extension for reply within third month	
1254 1,480	2254 740	Extension for reply within fourth month	
1255 2,010	2255 1,005	Extension for reply within fifth month	
1401 330	2401 165	Notice of Appeal	
1402 330	2402 165	Filing brief in support of an appeal	
1403 290	2403 145	Request for oral hearing	
1451 1,510	1451 1,510	Petition to institute a public use proceeding	
1452 110	2452 55	Petition to revive - unavoidable	
1453 1,330	2453 665	Petition to revive - unintentional	
1501 1,330	2501 665	Utility issue fee (or reissue)	
1502 480	2502 240	Design issue fee	
1503 640	2503 320	Plant issue fee	
1460 130	1460 130	Petitions to the Commissioner	
1807 50	1807 50	Processing fee under 37 CFR 1.17(q)	
1808 180	1808 180	Submission of Information Disclosure Stmt	
8021 40	8021 40	Recording each patent assignment per property (times number of properties)	
1809 770	2809 385	Filing a submission after final rejection (37 CFR 1.129(a))	
1810 770	2810 385	For each additional invention to be examined (37 CFR 1.129(b))	
1801 770	2801 385	Request for Continued Examination (RCE)	
1802 900	1802 900	Request for expedited examination of a design application	

Other fee (specify) _____

*Reduced by Basic Filing Fee Paid

SUBTOTAL (3) (\$)

(Complete if applicable)

SUBMITTED BY	Name (Print/Type)	Martin Fessenmaier	Registration No. (Attorney/Agent)	46697	Telephone 714-641-5100
Signature					Date December 23, 2003

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BIOCHIP CONFIGURATIONS AND METHODS

Field of The Invention

Analytic devices and methods, particularly as they relate to analytic devices and methods
5 using biochips.

Background of The Invention

Thousands of clinical tests are available for routine check-ups, diagnosis of conditions
and diseases, and many physicians rely heavily on the test results to establish a diagnosis and/or
treatment plan. Consequently, accuracy and reliability of such methods are of paramount
10 importance.

In numerous cases, calibration materials and controls are used as tools to assess accuracy
and precision of test results, and typically provide at least some information once a test protocol
has been executed. Recently, an active quality process control program for continuous control of
an analytic process with real-time automated error detection/correction, and documentation of
15 corrective actions was introduced (iQMTM on the GEM®Premier 3000). Such systems are often
reliable and provide accurate error reports, however, are typically difficult to integrate into
microarray-based assays. One of the major challenges in almost all microarray-based test systems
is the relatively strong variance of the system due to poor spotting quality, imprecise micro-
movements of motors or mechanical parts for scanning, and fluctuations in power, light sources
20 and/or detectors (*e.g.*, PMT).

Moreover, and especially where microarray systems are employed for nucleic acid
analysis, relatively expensive reagents and pre-amplification steps are often required to obtain a
reliable and quantifiable result from a test sample. For example, a number of primer extension
methodologies are known (see below) in the art, and are typically performed as single nucleotide
25 primer extension. However, such methods generally require expensive detectable dye-labeled
terminators such as acylo- or dideoxy nucleotides, and often need to be performed with pre-
amplified material.

Therefore, although there are numerous diagnostic devices and methods known in the art, there is still a need to provide improved biochip configurations and methods for analytic and diagnostic devices.

Detailed Description

5 The inventors discovered that numerous assays can be performed on a biochip with increased accuracy and simplicity using a biochip that includes a capture probe having an intrinsic reference marker, and or using a capture probe that serves as a substrate in a non-restricting enzymatic assay. While it is generally contemplated that the configurations and methods according to the inventive subject matter may be implemented with any biochip, 10 particularly preferred biochips are those described in our co-pending patent applications with the serial numbers 10/346879 (filed 1/17/03) and 10/433766 (filed 10/10/03), both of which are incorporated by reference herein.

Tagged Capture Probes

15 In one particularly preferred aspect of the inventive subject matter, a capture probe is tagged with a fluorophore, chemiluminophore, or otherwise detectable label, and the capture probe is immobilized in a predetermined position on the biochip. Thus, especially contemplated capture probes include antibodies, antigens, or nucleic acids (*e.g.*, DNA or RNA oligos). Typically, the capture is immobilized on a solid-phase (*e.g.*, glass, plastic or a membrane) or on/in a matrix.

20 It should be especially appreciated that the tagged capture probes can be used as internal quality control to check the quality and dimensions of the spots. Moreover, the tagged probes can also be exploited to design assays based on ratio determination. For example, in a sandwich immunoassay, the capture antibody can be tagged with the Cy3 dye while the reporter antibody is tagged with the Cy5 dye.

25 Therefore, an exemplary method of quantitative and/or qualitative analysis may include a step of tagging a capture probe with a detectable label. In another step, the tagged capture probe is immobilized on a biochip in a predetermined position (*e.g.*, on a solid-phase). In yet another step, unlabeled or labeled sample and/or a reporter probe is added to the tagged capture probe,

and hybridization or incubation of the sample is performed with the tagged capture probe and/or in the presence of the reporter probe. In still a further step, the labels of the tagged probes (and the reporter probes) are detected and quantified using suitable technologies (e.g., chemically, spectroscopically, or electrochemically).

5 In further especially preferred aspects, the label (*i.e.*, the tag) is at least one of a fluorophore, a chemiluminophore, and an electroactive group, while the tagged capture probe is preferably an antibody, an antigen, or an oligonucleotide. Where reporter probes are employed, the reporter probe is advantageously an antibody, an antigen, or an oligonucleotide. Of course, it should be recognized that depending on the chemical nature of the tag and/or the capture probe,
10 the tag can be covalently or non-covalently coupled to the probe. Furthermore, it should be appreciated that the tag on the tagged capture probe can be used during spotting for positional control, and/or control of the amount deposited onto the biochip. during spotting.

15 For additional control (*e.g.*, bleaching, quenching, identification, etc.) an additional tag can be immobilized without capture probe onto the biochip. Moreover, the tag on the capture probe may also be employed for quality control and ratio determination in an assay. Most preferably, spot morphology of the spot can be determined (*e.g.*, dimension of the spot).

Solid-Phase Primer Extension

20 The present inventive subject matter also relates to methods and reagents for determining the presence of a specific nucleotide in a specific position based on a microarray polynucleotide primer extension. In especially preferred aspects of such methods and reagents, the microarray polynucleotide primer extension does not require the use of dye-labeled chain terminators (*e.g.*, acylo-, or dideoxy nucleotides).

25 Primer extension as practiced herein is a result of polymerization of the complimentary deoxynucleotide and a detectable deoxynucleotide, and is not limited to the complimentary first base but proceeds up to the length of the available target and then terminates automatically. The primer extension based labeling without the use of terminators is an efficient way to label nucleic acids using microarray or solid-phase based technologies. Target specific hybridization is performed to achieve assay specificity.

Exemplary Protocol for on-chip non-limiting Primer Extension:

- (1) Prepare a DNA chip with specific capture oligonucleotides as shown in Figure 1.
- (2) Hybridize the immobilized capture oligonucleotides with 60 ul of hybridization solution (2.5 X SSC, 0.2% Tween 20, 12.5 mM HEPES, pH 7.5, 30 % Formamide) containing 5 CFTR template DNA and CB control DNA at 37 degree Celsius for 45 minutes.
- (3) After removing the hybridization solution, wash the chip with 100 ul of 1X SSC, 0.2% SDS one times and 0.5 X SSC solution twice at room temperature.
- (4) Apply primer extension solution 50 ul onto the chip and incubate at 37 degree for 30 min. (Primer extension solution: 0.5 uM of each dATP, dTTP and dGTP, 0.5 uM of Cy5 10 labeled dCTP, 1 unit of Terminator DNA polymerase and 1x reaction buffer).
- (5) Remove the reaction mixture, and wash the chip three times with 100 ul of 10 mM Tris HCl, 0.2 % Tween 20.
- (6) Excite the chip with lasers for Cy3 and Cy5 dyes and detect emission fluorescence 15 with a photomultiplier tube (PMT). The result using On-Chip Labeling using Primer Extension with Cy5 dCTP is shown in Figure 2.

Therefore, a method for determining a specific nucleotide at one or more location in a target nucleic acid may include one step in which a single stranded target nucleic acid is provided. In another step, the target nucleic acid is hybridized with a primer complimentary to the target nucleic acid, wherein the 3'-end of the primer binds to nucleotides flanking the specific 20 nucleotides at sites in the target nucleic acid. In still another step, the hybridized nucleic acid is exposed to a polymerase in a mixture containing at least one deoxynucleotide and one or more labeled deoxynucleotide.

In preferred aspects, the target nucleic acid is obtained from a human, animal, virus, bacteria, plant, or fungus (most preferably from a living specimen), and the above described 25 method is used for determination of specific nucleotide in predetermined position, species

identification, identification of diseases, predisposition to genetic diseases, fingerprinting, and/or pharmacogenomics.

Particularly preferred method will typically not require amplification of the target nucleic acid, but amplification is not excluded (*e.g.*, via polymerase chain reaction, LCR, etc. using 5 thermostable polymerase, ligase, or Klenow fragment.). The reaction mixture comprises at least one deoxynucleotide and a deoxynucleotide that is labeled for detection, and it is required that the primer hybridizes to the target single stranded nucleic acid. Detection is typically performed using fluorescence, chemiluminescence, colorimetrically, electrically or combination thereof (thus, the deoxynucleotide is labeled with fluorophore, chemiluminophore, enzyme, radioactive 10 or biotin, digoxin, or an electroactive label). Of course, it should be recognized that multiple fluorescent dyes can be used (*e.g.*, for determining a homozygote or a heterozygote, or for performing complex/multiplex analysis). Labeled or unlabeled deoxynucleotides can be used sequentially or simultaneously.

The following references (each of which is incorporated by reference herein) are provided 15 to reflect the level of general knowledge in the art. Basic Planning for Quality (Westgard QC, Inc.: 7614 Gray Fox Trail, Madison WI 53717). Basic QC Practices, Second Edition (Westgard QC, Inc.: 7614 Gray Fox Trail, Madison WI 53717), and U.S. Pat. Nos. 5846710, 6498012, 6503718, 6013431, 6479242, 5726015, 6355433, 6511803, and 6503718.

Thus, specific embodiments and applications of improved biochip configurations and 20 methods have been disclosed. It should be apparent, however, to those skilled in the art that many more modifications besides those already described are possible without departing from the inventive concepts herein. The inventive subject matter, therefore, is not to be restricted except in the spirit of the present disclosure. Moreover, in interpreting the specification, all terms should be interpreted in the broadest possible manner consistent with the context. In particular, the terms 25 "comprises" and "comprising" should be interpreted as referring to elements, components, or steps in a non-exclusive manner, indicating that the referenced elements, components, or steps may be present, or utilized, or combined with other elements, components, or steps that are not expressly referenced.

Cy5 positive	→	○ ○ ○ ○ ○ ○ ○ ○	←	Buffer only
MDR 1 capture Probe (30 mer)	→	● ● ● ● ● ● ● ●	←	CFTR capture Probe (30 mer)
MDR 1 capture Probe (21 mer)	→	● ● ● ● ● ● ● ●	←	CFTR capture Probe (20 mer)
Cy5 positive	→	● ● ● ● ● ● ● ●	←	CB 024 control
Buffer only	→	● ● ● ● ● ● ● ●	←	CB 828 control

Figure 1

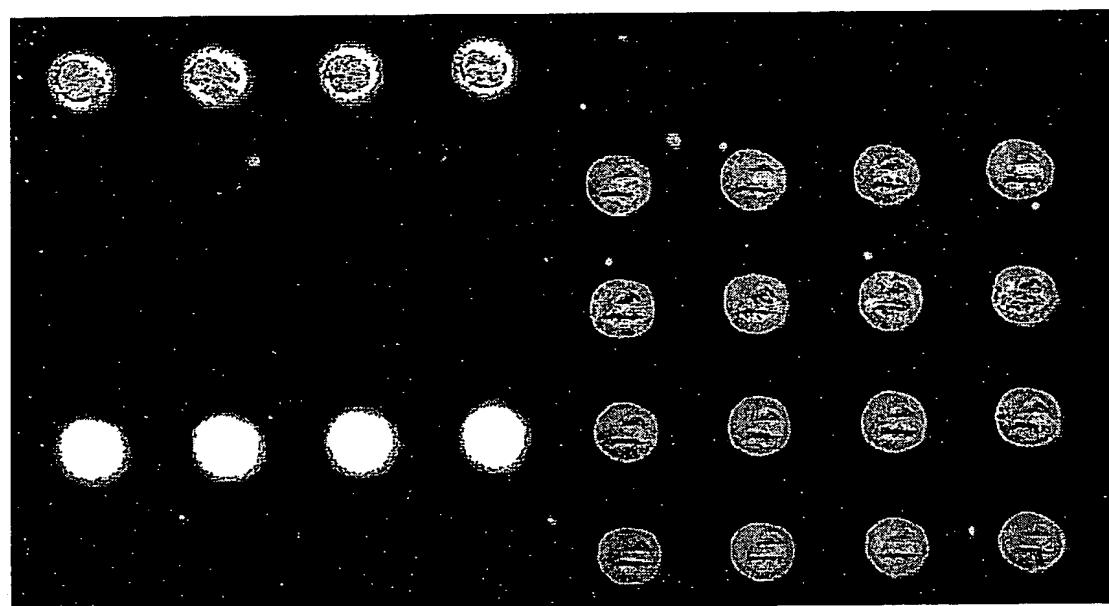


Figure 2

Document made available under the Patent Cooperation Treaty (PCT)

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